

BAR One Step Barbiturates Test Device (Urine) Package Insert

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For professional in vitro diagnostic use only.

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INTENDED USE

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The BAR One Step Barbiturates Test Device (Urine) is a lateral flow chromatographic immunoassay for the detection of Barbiturates in urine at a cut-off concentration of 300 ng/mL. This test will detect other related compounds, please refer to the Analytical Specificity table in this package insert.

This assay provides only a qualitative, preliminary analytical test result. A more specific alternate chemical method must be used in order to obtain a confirmed analytical result. Gas chromatography/mass spectrometry (GC/MS) is the preferred confirmatory method. Clinical consideration and professional judgment should be applied to any drug of abuse test result, particularly when preliminary positive results are used.

INTRODUCTION

Barbiturates are central nervous system depressants. They are used therapeutically as sedatives, hypnotics and anticonvulsants. Barbiturates are almost always taken orally as capsules or tablets. The effects resemble those of intoxication with alcohol. Chronic use of barbiturates leads to tolerance and physical dependence.

Short acting Barbiturates taken at 400 mg/day for 2-3 months produces a clinically significant degree of physical dependence. Withdrawal symptoms experienced during periods of drug abstinence can be severe enough to cause death.

Only a small amounts (less than 5%) of most Barbiturates are excreted unaltered in the urine.

The approximate detection time limits for Barbiturates are:

Short acting(e.g. Secobarbital)	100mg PO (oral)	4.5days
Long acting(e.g. Phenobarbital)	400mg PO (oral)	7days

PRINCIPLE

The BAR One Step Barbiturates Test Device (Urine) is an immunoassay based on the principle of competitive binding. Drugs that may be present in the urine specimen compete against the drug conjugate for binding sites on the antibody.

During testing, a urine specimen migrates upward by capillary action. Barbiturates, if present in the urine specimen below the cut-off level, will not saturate the binding sites of the antibody in the test. The antibody coated particles will then be captured by immobilized Barbiturates, protein conjugate and a visible colored line will show up in the test line region. The colored line will not form in the test line region if the Barbiturates level exceeds the cut-off level, because it will saturate all the binding sites of anti-Barbiturates antibodies.

A drug-positive urine specimen will not generate a colored line in the test line region because of drug competition, while a drug-negative urine specimen or a specimen containing a drug concentration less than the cutoff will generate a line in the test line region. To serve as a procedural control, a colored line will always appear at the control line region indicating that proper volume of specimen has been added and membrane wicking has occurred.

REAGENTS

The test contains drug-bovine protein antigen conjugate on the membrane and the conjugate pad of each test contains monoclonal anti-

drug antibody.

KIT COMPONENTS

Individually packed Test	Each Device contains a strip with			
Devices	colored conjugates and reactive			
	reagents pre-spreaded at the			
	corresponding regions.			
Package insert	For operation instruction.			
MATERIALS REQUIRED BUT NOT PROVIDED				

MATERIALS REQUIRED BUT NOT PROVIDED

Specimen collection	For specimens collection use.
container	
Timer	For timing use.

PRECAUTIONS

- · For professional in vitro diagnostic use only.
- Do not use after expiration date indicated on the package. Do not use the test if its foil pouch is damaged. Do not reuse tests.
- This kit contains products of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not totally guarantee the absence of transmissible pathogenic agents. It is therefore, recommended that these products be treated as potentially infectious, and handled observing the usual safety precautions (do not ingest or inhale).
- Avoid cross-contamination of specimens by using a new specimen collection container for each specimen obtained.
- Read the entire procedure carefully prior to performing any tests.
- Do not eat, drink or smoke in the area where the specimens and kits are handled. Handle all specimens as if they contain infectious agents. Observe established precautions against microbiological hazards throughout the procedure and follow the standard procedures for proper disposal of specimens. Wear protective clothing such as laboratory coats, disposable gloves and eye protection when specimens are assayed.
- · Humidity and temperature can adversely affect results.
- The used testing materials should be discarded in accordance with local, state and/or federal regulations.

STORAGE AND STABILITY

- The kit should be stored at 2-30°C until the expiry date printed on the sealed pouch.
- The test must remain in the sealed pouch until use.
- Do not freeze.
- Cares should be taken to protect components in this kit from contamination. Do not use if there is evidence of microbial contamination or precipitation. Biological contamination of dispensing equipment, containers or reagents can lead to false results.

SPECIMEN COLLECTION AND STORAGE

- The urine specimen must be collected in a clean and dry container. Urine collected at any time of the day may be used. Urine specimens exhibiting visible particles should be centrifuged, filtered, or allowed to settle to obtain clear specimen for testing.
- Collected urine specimens must be put in clear and dry containers.
- Perform the testing immediately after the specimen collection. Do not leave the specimens at room temperature for prolonged periods. Specimens may be stored at 2-8°C for up to 48 hours. For long term storage, specimens should be kept below -20°C.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Avoid repeated freezing and thawing of specimens.
- Pack the specimens in compliance with applicable regulations for transportation of etiological agents, in case they need to be shipped.
 PROCEDURE

Bring tests, specimens and/or controls to room temperature (15-30°C) before use.

- 1. Bring the pouch to room temperature before opening it. Remove the test device from the sealed pouch and use it as soon as possible.
- 2. Place the test device on a clean and level surface. Hold the dropper vertically and **transfer 3 full drops of urine** (approx. 100 μ L) to the specimen well (S) of the test device, and then start the timer. Avoid trapping air bubbles in the specimen well (S).
- 3. Wait for the colored line(s) to appear. **Read results at 5 minutes.** Do not interpret the result after 10 minutes.

INTERPRETATION OF RESULTS

POSITIVE RESULT:



Only one colored band appears in the control region (C). No apparent colored band appears in the test region (T).

NEGATIVE RESULT:



Two colored bands appear on the membrane. One band appears in the control region (C) and another band appears in the test region (T).



Control band fails to appear. Results from any test which has not produced a control band at the specified reading time must be discarded. Please review the procedure and repeat with a new test. If the problem persists, discontinue using the kit immediately and contact your local distributor.

NOTE:

- The intensity of the color in test region (T) may vary depending on the concentration of aimed substances present in the specimen. Therefore, any shade of color in the test region should be considered negative. Besides, the concentration level can not be determined by this qualitative test.
- 2. Insufficient specimen volume, incorrect operation procedure, or performing expired tests are the most likely reasons for control band failure.

QUALITY CONTROL

- Internal procedural controls are included in the test. A colored band appearing in the control region (C) is considered an internal positive procedural control. It confirms sufficient specimen volume and correct procedural technique.
- External controls are not supplied with this kit. It is recommended that positive and negative controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance.

LIMITATIONS OF THE TEST

- 1. The BAR One Step Barbiturates Test Device (Urine) provides only a qualitative, preliminary analytical result. A secondary analytical method must be used to obtain a confirmed result. Gas chromatography/mass spectrometry (GC/MS) is the preferred confirmatory method.^{1,2}
- 2. It is possible that technical or procedural errors, as well as other interfering substances in the urine specimen may cause erroneous results.
- 3. Adulterants, such as bleach and/or alum, in urine specimens may produce erroneous results regardless of the analytical method used. If adulteration is suspected, the test should be repeated with another urine specimen.
- 4. A positive result indicates presence of the drug or its metabolites but does not indicate level of intoxication, administration route or concentration in urine.
- 5. A negative result may not necessarily indicate drug-free urine. Negative results can be obtained when drug is present but below the cut-off level of the test.

6. Test does not distinguish between drugs of abuse and certain medications.

PERFORMANCE CHARACTERISTICS

A. Accuracy

123 clinical urine specimens were analyzed by GC-MS and by the BAR One Step Barbiturates Test Device (Urine).Each test was performed by three operators. Samples were divided by concentration into five categories: negative, less than half the cutoff, near cutoff negative, near cutoff positive, and high positive. Results were as follows:

The Barl	BAR One Step Diturates t Device	Neg	Neg. (< – 50% cutoff)	GO Near cutoff neg. (-50% cutoff to	C/MS Near cutoff pos. (cutoff to +50%	Pos. (> +50 % cutoff	% agreem ent with GC/MS
	D 111	-	-	cutoff)	cutoff))	,
BAR	Positive	0	0	0	11	50	95.31%
300	Negative	38	12	9	3	0	100%

B. Precision

A study was conducted at three physician offices for Barbiturates(300 ng/mL)by professional operators using three different lots of product to demonstrate the within run, between run and between operator precision. An identical panel of coded specimens, containing drugs at the concentration of \pm 50% and \pm 25% cut-off level, was labeled as a blind and tested at each site. The results are given below:

Drug Conc.	n	Site A		Site B		Site C	
Di ug Colic.	per site	-	+	-	+	-	+
Negative	10	10	0	10	0	10	0
-50% Cut-off	10	10	0	10	0	10	0
-25% Cut-off	10	9	1	10	0	9	1
+25% Cut-off	10	1	9	1	9	0	10
+50% Cut-off	10	0	10	0	10	0	10

C. Effect of Urinary Specific Gravity

Fifteen (15) urine samples of normal, high, and low specific gravity ranges (1.000-1.037) were spiked with drugs at 50% below and 50% above cut-off levels respectively. The BAR One Step Barbiturates Test Device (Urine) was tested in duplicate using fifteen drug free urine and spiked urine samples. The results demonstrate that varying ranges of urinary specific gravity do not affect the test results.

D. Effect of Urinary pH

The pH of an aliquoted negative urine pool was adjusted to a pH range of 5 to 9 in 1 pH unit increments and spiked with drugs at 50% below and 50% above cut-off levels. The spiked, pH adjusted urine was tested with the BAR One Step Barbiturates Test Device (Urine). The results demonstrate that varying ranges of pH do not interfere with the performance of the test.

E. Cross-Reactivity

The following tables list the concentrations of compounds (ng/mL) above which the BAR One Step Barbiturates Test Device (Urine) identified positive results at 5 minutes.

Barbiturates related Compound	Concentration (ng/mL <u>)</u>	
Amobarbital	300	
Aprobarbital	200	
Barbital	300	
Butabarbital	75	

Butalbital	2500
Pentobarbital	300
Phenobarbital	100

F. Non Cross-Reacting Compounds

The following compounds yielded negative results up to a concentration of 100 μ g/mL:

Penfluridol

salt

salt

Penicillin G potassium

Penicillin G sodium

Phenelzine Sulfate

Perphenazine

Phenothiazine 2-Phenylethylamine

Pioglitazone

Piracetam Pravastatin sodium

Prednisone

Promethazine

hvdrochlorine

Pvridoxine

Pyrogallic

Quinine Ouinolinic acid

R,R(-)-

Ranitidine

Riboflavin

Rifampicin

Risperidone

Salicvlic acid

Simvastatin

Sodium 2-

Sulindac

Thiamine

Tolbutamide Topiramate 2,4,7-Triamino-6-Phenylpteridine Trimethoprim Tryptamine Tyramine Uric acid

Tetracycline

Sertraline HCl

Propylvalerate

Sulfamethazine

Tetrahydrozoline Theophylline

Thioridazine solution

6-Propyl-2-thiouracil

Pyrilamine Maleate

Quetiapine Fumarate

Pseudoephedrine Ranitidine base

Procaine

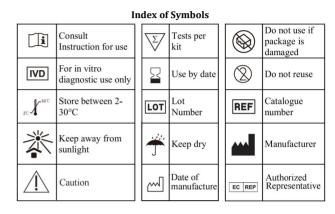
Phenacetin

4-Acetamidophenol	Gatifloxacin
Acetaminophen	Gemfibrozil
Acetylsalicylic Acid	Gentisic Acid
Albumin Amoxicillin Ampicillin trihydrate Aspartame Atropine Baclofen Benzoic Acid Berberine Chloride Hydrate	Gliclazide Glipizide Glyburide Guaiacol Guaifenesin Hemoglobin Hydralazine HCl Hydrochlorothiazide Hydrocortisone
Bilirubin	Ibuprofen
Caffeine	Isoprenaline
Cephalexin Cephradine Chloral hydrate Chloramphenicol Chlorpheniramine Maleate Chlorpromazine	Ketoconazole Ketoprofen Lamotrigine L-Ascorbic acid Levofloxacin Lidocaine
Cholesterol	Lidocaine Monohydrate
Ciprofloxacin hydrate	Lisinopril Dihydrate
Clarithromycin Clonidine solution Creatinine D(-)-Norgestrel d,l-Propranolol Deoxycorticosterone Dextromethorphan solution	Lithium carbonate Loperamide Loratadine L-Thyroxine sodium Maprotiline Meprobamate Minocycline
Diciofenac	Mosapride Citrate
Diflunisal	Nalidixic acid
Digoxin	Naloxone HCl
4-Dimethyl- aminoantipyrine	Naltrexone HCl
Diphenhydramine 5,5-Diphenylhydantoin D-Lactose monohydrate D-Leucyl-L-tyrosine	Naproxen Nicotinamide Nicotinic acid Nifedipine
Hydrate Dopamine	Nimodipine
Droperidol Enalapril Maleate	Norethisterone Acetate Norfloxacin Nicotinic
Erythromycin	Noscapine
Estradiol Estrone Ethyl 4-aminobenzoate Fluoxetine	(±)-Octopamine Ofloxacin Olanzapine Oxalic acid, anhydrous

Fotemustine	Oxolinic acid	(±)-Verapamil
Furosemide	Paliperidone	Vitamin B1
Gabapentin	Pantoprazole sodium	Zomepirac

LITERATURE REFERENCES

- 1. Baselt RC. <u>Disposition of Toxic Drugs and Chemicals in Man.</u> 2nd Ed. Biomedical Publ., Davis, CA. 1982; 488
- 2. Hawks RL, CN Chiang. Urine Testing for Drugs of Abuse. National Institute for Drug Abuse (NIDA), Research Monograph 73, 1986



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